SHAVINGS REFERENCE

* Shavings are also referred to as curls or scrolls.

The following information is to help the requester understand the scope of the technique the RHCL uses to obtain shavings.

**Procedures:** The investigator must submit a request via the iLab system for shavings. The thickness for shavings most commonly requested from investigators is 10 microns, but size can vary based on the requester’s requirements (up to 100 microns).

*Please note: For any shavings set to be used for RNA extraction or molecular analysis at a later time, we highly recommend that service of DNA/RNA prep be requested alongside the cutting of shavings in order to prevent contamination.*

**Scope:**

**Paraffin Samples:**

1. The microtome station (including microtome and sectioning instruments) will be cleaned with disinfectants and treated with DNA/RNase decontaminant solution (*if prep is requested*).
2. Eppendorf tubes will only be handled with gloves throughout the entire preparations and sectioning.
3. In order to prevent cross-contamination between samples, the area of the blade used for cutting will be changed for each sample.
4. Shavings will be produced at the specified thickness to generate curls that will be placed in sterile Eppendorf tubes, labeled with the sample ID.

**Frozen Samples:**

1. When the lab receives frozen samples, they will be stored at -20°C (*or -80°C if requested*) until sectioning.
2. Cryostat and cryotomy tools will be thoroughly cleaned and treated with DNA/RNase decontaminant solution (*if prep is requested*).
3. Cryostats are routinely maintained at approximately -20°C. Therefore, frozen samples will be equilibrated to -20°C before sectioning begins.
4. Eppendorf tubes will only be handled with gloves throughout the entire preparations and sectioning.
5. In order to prevent cross-contamination between samples, the area of the microtome blade will be changed for each sample.
6. Shavings will be produced at the specified thickness to generate curls that will be placed in sterile Eppendorf tubes, labeled with the sample ID.
7. At the time of checkout, it is recommended that samples be transported in a manner that insulates frozen temperatures (*i.e. dry ice*).

**If there are any questions regarding the services offered, please contact the Research Histology Core Lab at (713) 792-3119 or via email at RHFCoreLab@mdanderson.org.**